

Using semiochemicals to manipulate populations of *Diorhabda* spp for wildlife management.

11/12/2020

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Semiochemicals,
Lovelock 2004

“Science is ahead of practice” Peter Skidmore, Walton Family Foundation
Environment Program, REW Conference, 2-04-20



Questions (to complement restoration efforts)

1. Which beetles will be in the system?

Bioinformatics

2. How often and when will they defoliate?

Phenology and phenotype tracking

3. How can we manipulate populations?

Semiochemicals to modify beetle behavior



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Monitoring and manipulation of *Diorhabda* populations within southwestern willow flycatcher nesting territories

Alex Gaffke, Tom Dudley, Dan Bean

This proposed work will enable *Diorhabda* populations to be accurately monitored and future distributions predicted as they move into areas where southwestern willow flycatchers (SWFL) are known to nest in tamarisk. Information obtained through monitoring can inform efforts to manipulate spatial distribution of populations through the use of behaviorally active compounds (semiochemicals).

A more focused consideration of how the project can be applied in the upper Gila River watershed, and also our responses to questions from the US Fish & Wildlife Service, are included as appendices at the end of this document. The proposed work will be divided into three sections:

- 1) using semiochemicals to manipulate *Diorhabda* populations on a local spatial scale;
- 2) using developmental data to predict *Diorhabda* distribution and colonization events;
- 3) using molecular genetic data to identify *Diorhabda* populations, species and their hybrids.



Monitoring and manipulation of *Diorhabda* populations within southwestern willow flycatcher nesting territories

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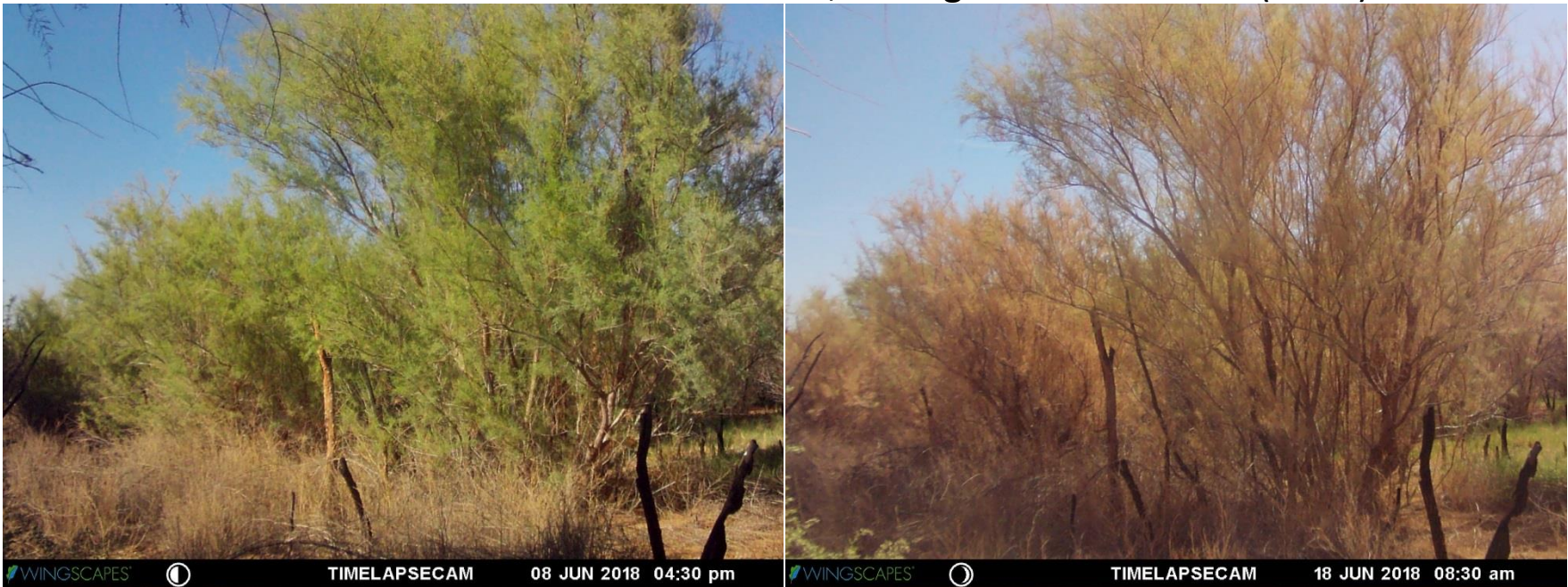


Beetles in the genus *Diorhabda* form aggregations as part of their behavioral strategy





Defoliation at the Cibola NWR, third generation larvae (2019)



1. Adults emerge from the leaf litter and feed
2. Adults aggregate on larva-free trees
3. They mate and lay eggs
4. Adults move on, larvae feed and defoliate
5. Mature larvae drop to the ground and pupate
6. The next generation adults emerge from the leaf litter

Defoliation at the Cibola NWR, third generation larvae



Overwintered spring adults

1. Adults emerge from the leaf litter and feed
2. Adults aggregate on larva-free trees
3. They mate and lay eggs
4. Adults move on, larvae feed and defoliate
5. Mature larvae drop to the ground and pupate
6. The next generation adults emerge from the leaf litter

Defoliation at the Cibola NWR, third generation larvae



Aggregating reproductive adults

1. Adults emerge from the leaf litter and feed
2. Adults aggregate on larva-free trees
3. They mate and lay eggs
4. Adults move on, larvae feed and defoliate
5. Mature larvae drop to the ground and pupate
6. The next generation adults emerge from the leaf litter

Defoliation at the Cibola NWR, third generation larvae



Reproductive adults avoid these trees



Defoliating late-stage larvae

1. Adults emerge from the leaf litter and feed
2. Adults aggregate on larva-free trees
3. They mate and lay eggs
4. Adults move on, larvae feed and defoliate
5. Mature larvae drop to the ground and pupate
6. The next generation adults emerge from the leaf litter

Reproductive adults avoid these trees



Pushed away from trees with heavy larval feeding



Pulled toward reproductive adult swarms

Nature's "push/pull" helps determine local beetle densities and the level and timing of defoliation

1. Adults emerge from the leaf litter and feed
2. Adults aggregate on larva-free trees ←
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Pushed away from trees with heavy larval feeding



Pulled toward reproductive adult swarms

What chemical signals mediate these behaviors?

Nature's "push/pull" helps determine local beetle densities and the level and timing of defoliation

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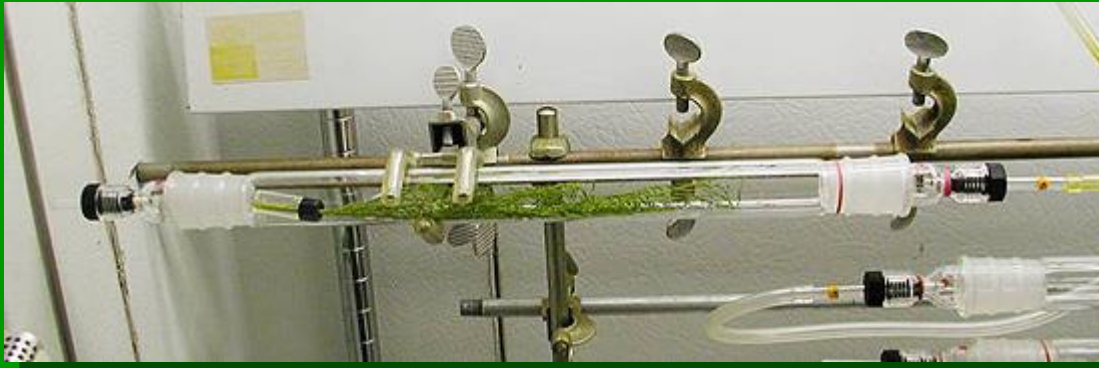
Allard Cossé checks field trials of attractants
Lovelock, NV, 2004



Bob Bartelt monitors
pheromone-baited trap,
Lovelock, 2003

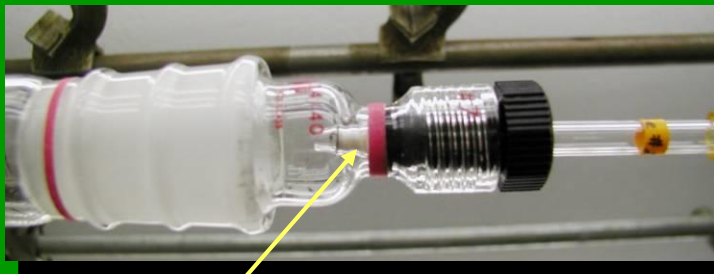
A pheromone blend produced by reproductive, feeding males mediates the formation of reproductive swarms of adult beetles, attracting females as well as other males

Collection of volatiles from tamarisk and feeding beetles



Collector tube with foliage and beetles

Beetles on foliage

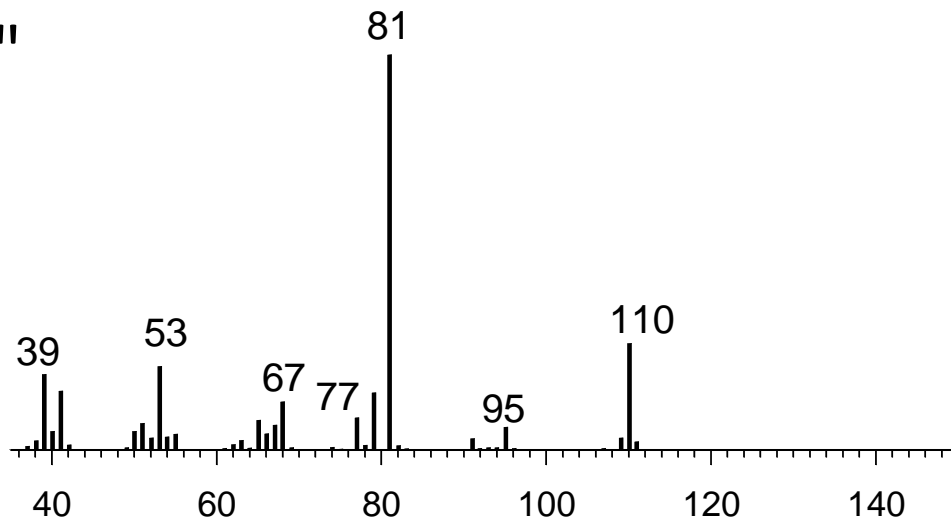


Close up of Super-Q filter

- Draw volatiles emitted from feeding beetles into filter of porous polymer ("Super-Q") with gentle vacuum; later on, rinse filter with solvent.
- On the plus side: Beetles + food is a "natural" situation; good chance of pheromone emission.
- On the minus side: plant compounds will also be collected.

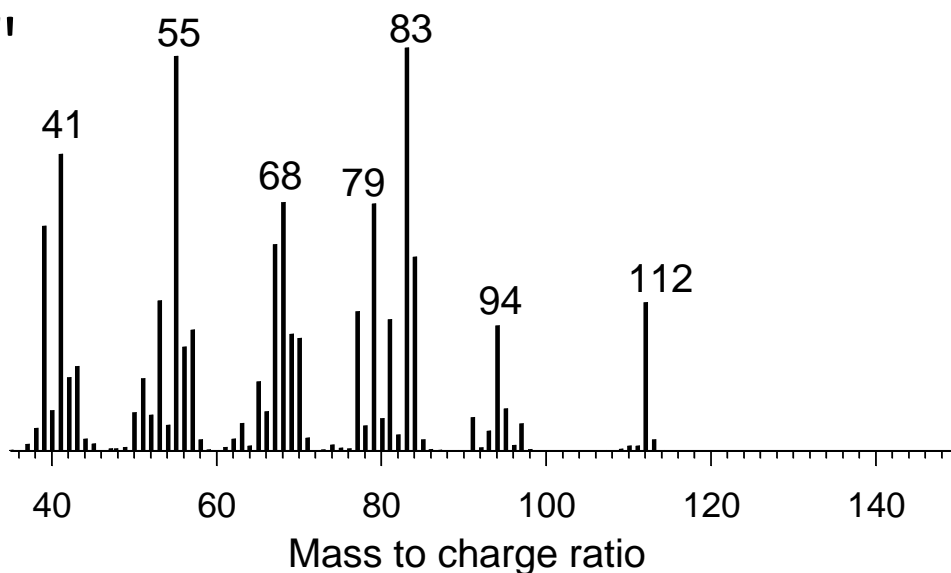
Mass spectra of male-specific compounds and ID's, based on MS library and analytical comparison with standards

"A"



(2E,4Z)-2,4-heptadienal
= "2E,4Z-7:Ald"

"B"



(2E,4Z)-2,4-heptadien-1-ol
= "2E,4Z-7:OH"

Cossé et al., 2005, J. Chem.
Ecol.



Allard Cossé checks field trials of attractants
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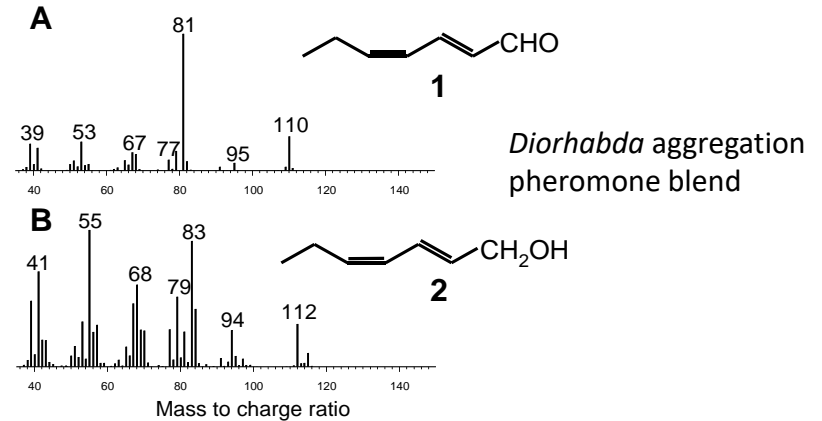
Bob Bartelt monitors
pheromone-baited trap,
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Discovery of the male-emitted aggregation-pheromone blend and plant-based attractants fueled efforts to use semiochemicals to:

1. Monitor *Diorhabda* populations at low densities
2. Assist in establishment of *Diorhabda* by keeping the adults in one place
3. Manipulate beetle populations with wildlife management as a goal ←



Alex Gaffke, Montana State University
 Beetle herding using behaviorally active
 compounds (semiochemicals) in *Diorhabda*



Gaffke, A. M., S. E. Sing, T. L. Dudley, D. W. Bean, J. A. Russak, A. Mafra-Neto, P. A. Grieco, R. K. D. Peterson, and D. K. Weaver. 2018. Semiochemicals to enhance herbivory by *Diorhabda carinulata* aggregations in saltcedar (*Tamarix* spp.) infestations. *Pest Management Science* 74(6): 1494 -1503.

Gaffke, A. M., S. E. Sing, T. L. Dudley, D. W. Bean, J. A. Russak, A. Mafra-Neto, P. A. Grieco, R. K. D. Peterson, and D. K. Weaver. 2019. Field demonstration of a semiochemical treatment that enhances *Diorhabda carinulata* biological control of *Tamarix* spp. *Scientific Reports* 9: 1305 <https://doi.org/10.1038/s41598-019-49459-5>.(53)

Gaffke AM, Sing SE, Dudley TL, Bean DW, Russak JA, Mafra-Neto A, Peterson RKD, Weaver DK (2020). Establishing *Diorhabda carinulata*: Impact of release disturbances on pheromone emission and influence of pheromone lures on establishment. *Journal of Chemical Ecology* 46: 378-386





Pheromone
treatment
2013

Untreated
Plants with
no impacts
from
Diorhabda



Impacts of
pheromone
treatment:
high levels
of dieback



Alex Gaffke
Beetle herding using behaviorally active
compounds (semiochemicals) in *Diorhabda*

Discovery of compounds repellent to
Diorhabda enable a “Push/Pull” strategy for
spatial manipulation of populations

Gaffke AM, Sing SE, Miller JG, Dudley TL, Bean DW, Peterson RKD, Weaver DK (2020). An herbivore-induced plant volatile from saltcedar (*Tamarix* spp.) is repellent to *Diorhabda carinulata* (Coleoptera: Chrysomelidae). *Environmental Entomology*
[https://doi: 10.1093/ee/nvaa079](https://doi.org/10.1093/ee/nvaa079)

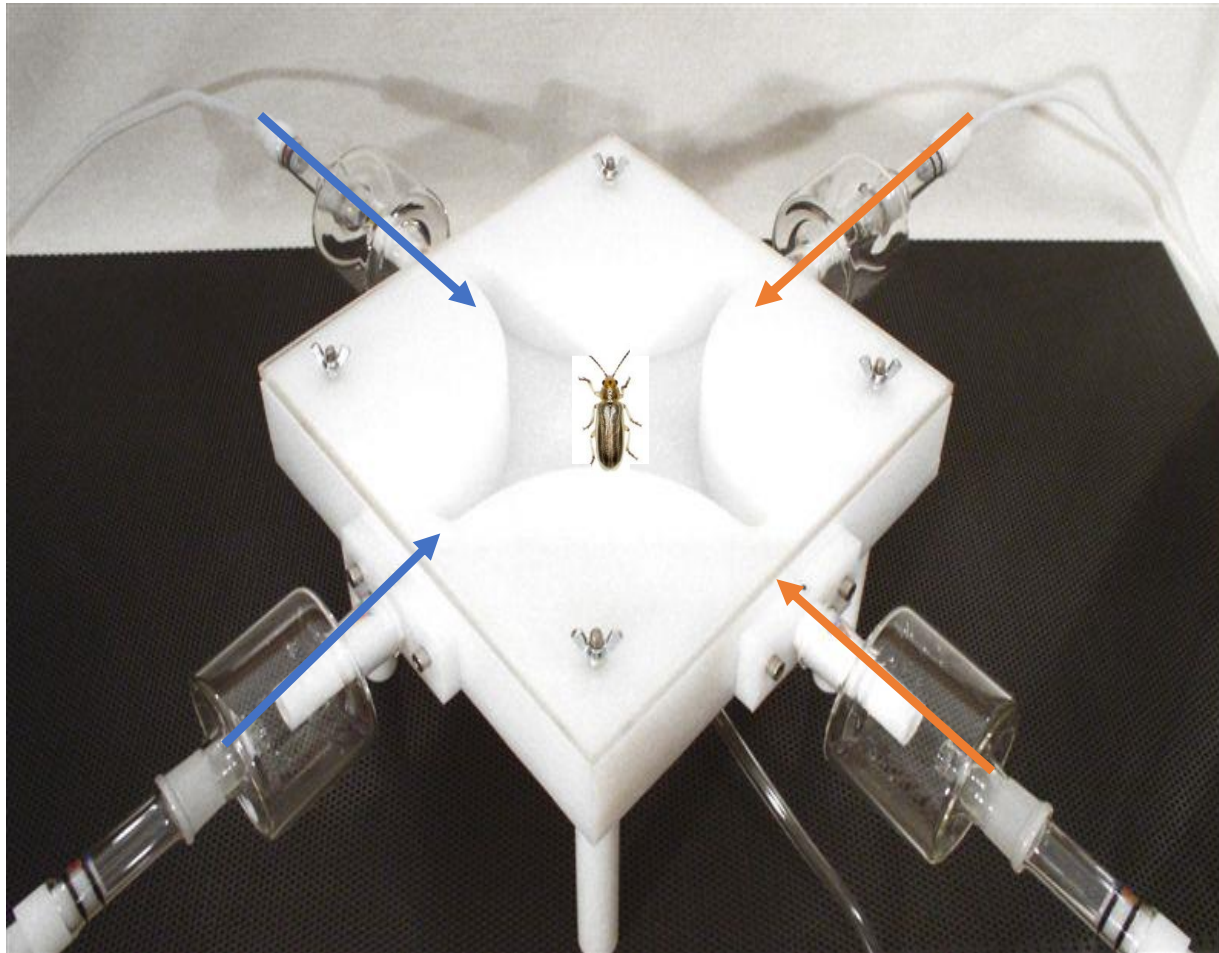
Push away from critical
nesting habitat

Pull toward expendable
tamarisk stands

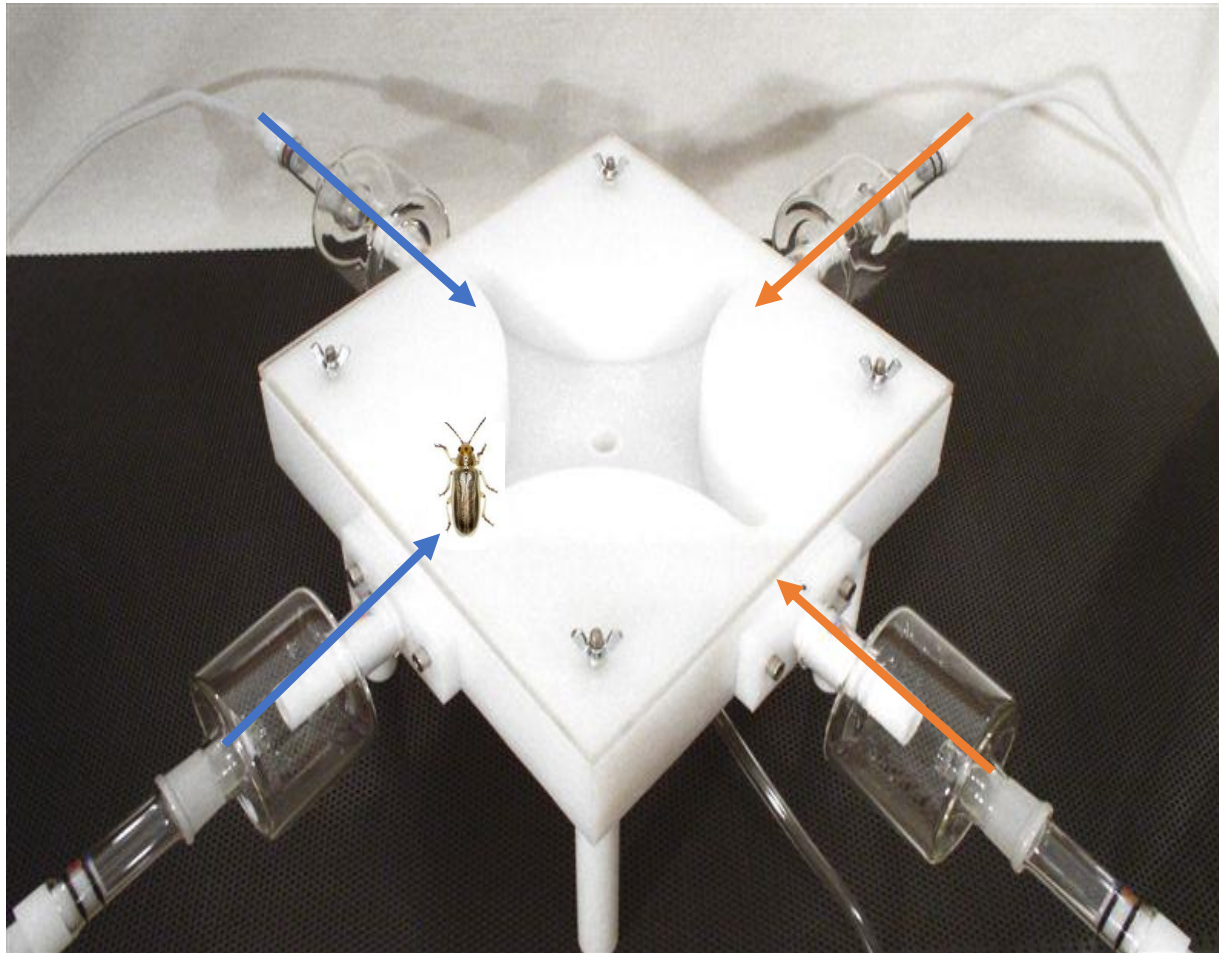
Quantities of **4-oxo-(E)-2-hexanal** released from *Tamarix* foliage plus adult *D. carinulata*, beetles alone and mechanically damaged foliage as controls

Emission rate			
Component	Mean (ng/beetle/day)	Standard Error	<i>N</i>
Adult males	0.70	± 0.20	10
Adult females	2.63	± 1.10	10
Control foliage	0.0	± 0.0	4
Mechanically damaged foliage	0.0	± 0.0	4
Adults without foliage	0.0	± 0.0	4

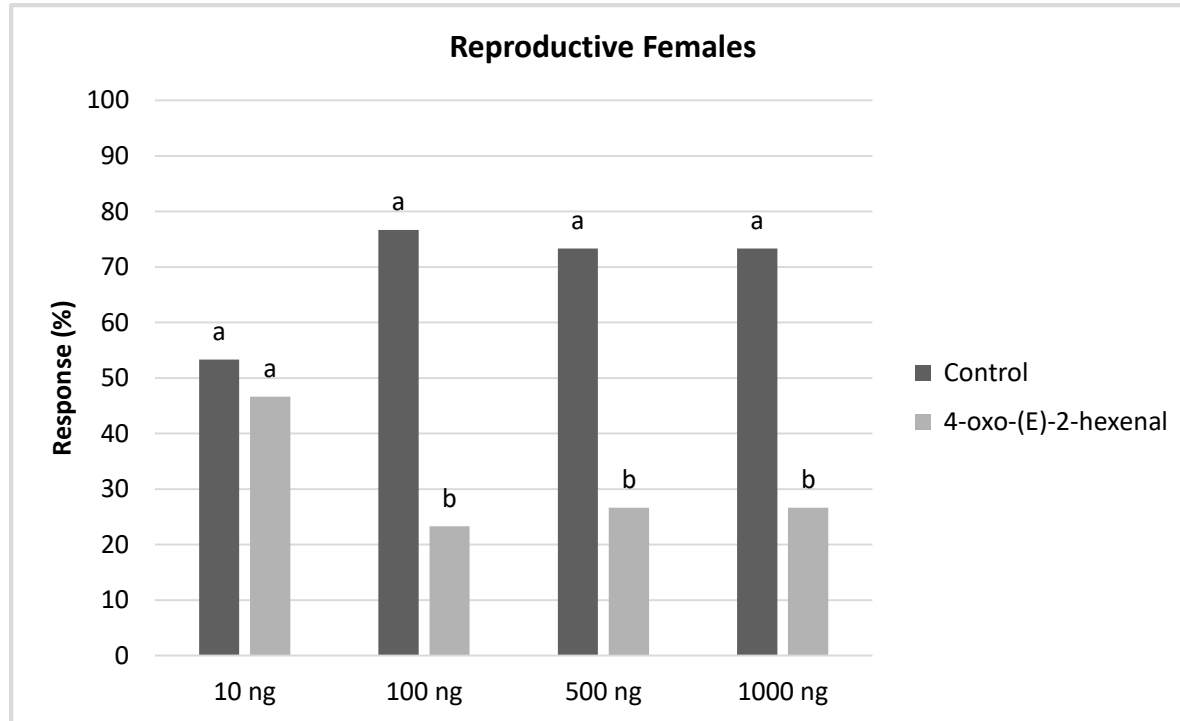
Behavioral bioassay



Behavioral bioassay



Repellent compounds are currently under investigation



Repellent Field Trials

Location	Date	Mean Adult Capture \pm SE		
		Control	Treatment	<i>P</i>
Saltcreek, CO	07/14/2018	23.1 \pm 8.5	9.6 \pm 3.1	0.04
Rangely, CO	7/31/2018	0.36 \pm 0.2	0.52 \pm 0.2	0.5
Cheney, CO	08/04/2018	1.7 \pm 0.6	1.0 \pm 0.3	0.2
Blythe, CA	08/20/2018	5.0 \pm 1.0	1.9 \pm 0.6	0.04
Blythe, CA	08/27/2018	1.4 \pm 1.1	1.6 \pm 0.4	0.35

Relevant points from Gaffke's work

1. A system was developed, in cooperation with a private company, for extended release of the male emitted pheromone (the system uses SPLAT, a proprietary slow-release formulation based on a waxy matrix that enables slow vaporization of the compounds).
2. Beetles can be herded to areas where they will defoliate tamarisk, while being distracted from other nearby trees. A compound was also found which repels female beetles and may prove useful as the push portion of a push-pull strategy
3. This work can be of immediate use to resource managers, providing a stop-gap measure to protect nesting birds from defoliation events while restoration is underway.

SPLAT pheromone release system



Implementation

1. Synthesize pheromone, develop strategic plan for the timing and spatial arrangement of lure placement
2. Work closely with those striving to restore the Gila watershed and increase ecosystem services (Gila Watershed Partnership and others)
3. Increase community education and involvement by providing the Gila Watershed Partnership with educational materials
4. Work closely with those involved in monitoring southwestern willow flycatcher populations and those involved in restoration of critical habitat
5. Coordinate with scientists working on various aspects of *Diorhabda* biology in order to better understand and predict life cycles and behavior of the beetle



Catch and kill



Pull with attractants



Push with repellents



Early season use traps to catch and kill adults early in the log phase of population growth



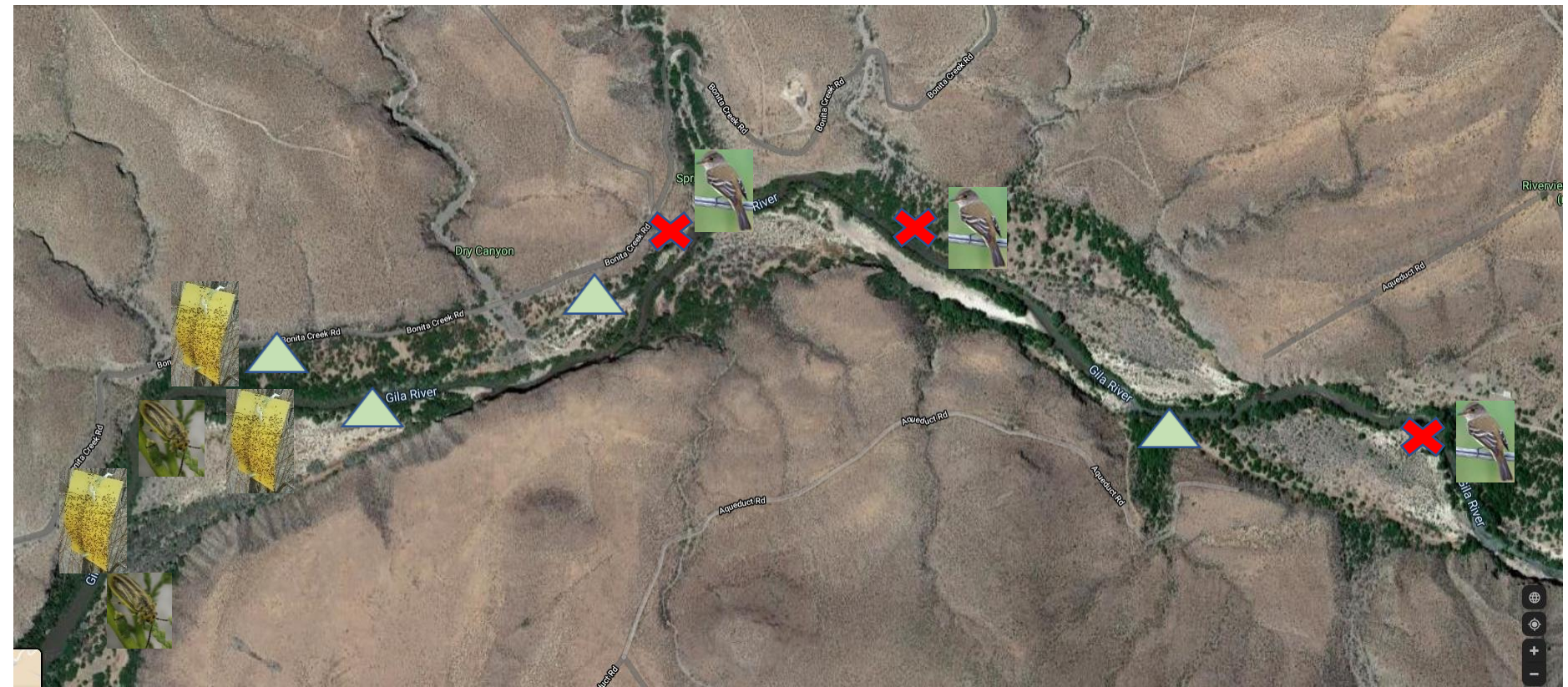
Catch and kill



Pull with attractants



Push with repellents



Later in the season use attractants to distract beetles from nesting territories of the SWFL. Use repellents to discourage beetles from colonizing near SWFL nests.



Catch and kill



Pull with attractants



Push with repellents

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Which beetles are in the upper Gila drainage?

Diorhabda sublineata



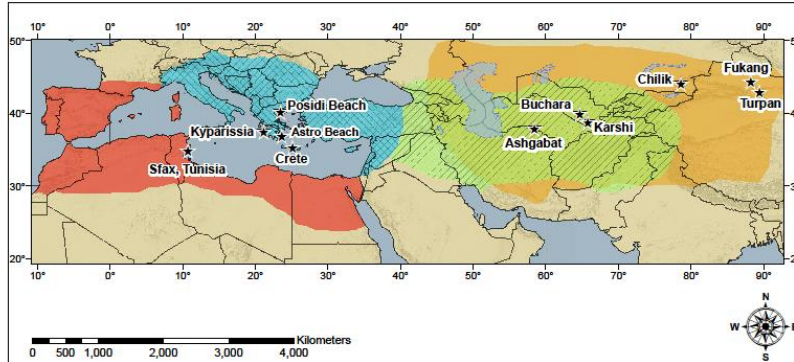
Diorhabda elongata



Diorhabda carinata



Diorhabda carinulata



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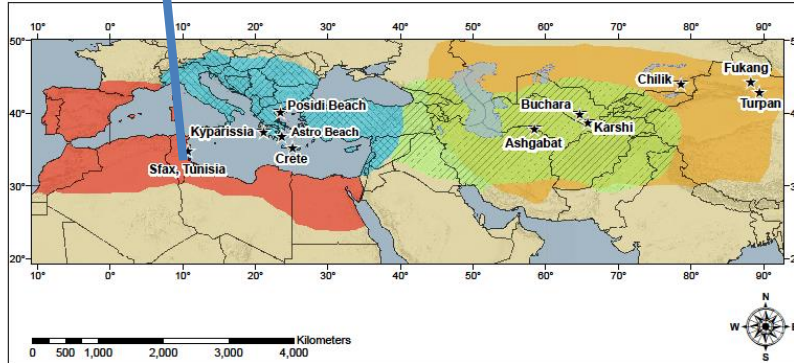
Diorhabda elongata



Diorhabda carinata



Diorhabda carinulata



Their great great...great great grandmothers came from Tunisia

personal communication,
Zeynep Özsoy, Colorado Mesa
University, based on
mitochondrial CO1 DNA
sequence

Which beetles are in the upper Gila drainage?

Diorhabda sublineata



X

Diorhabda elongata



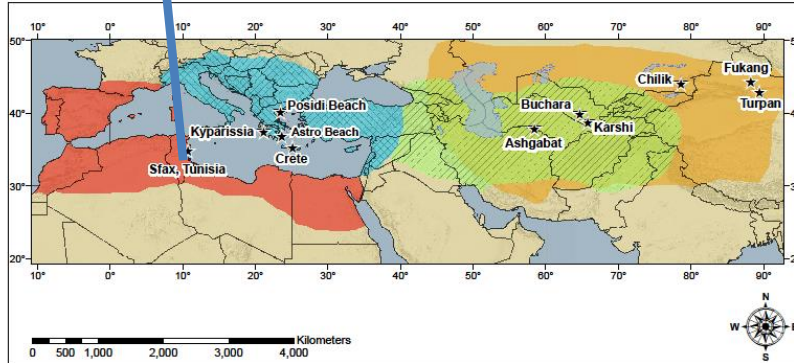
X

Diorhabda carinata



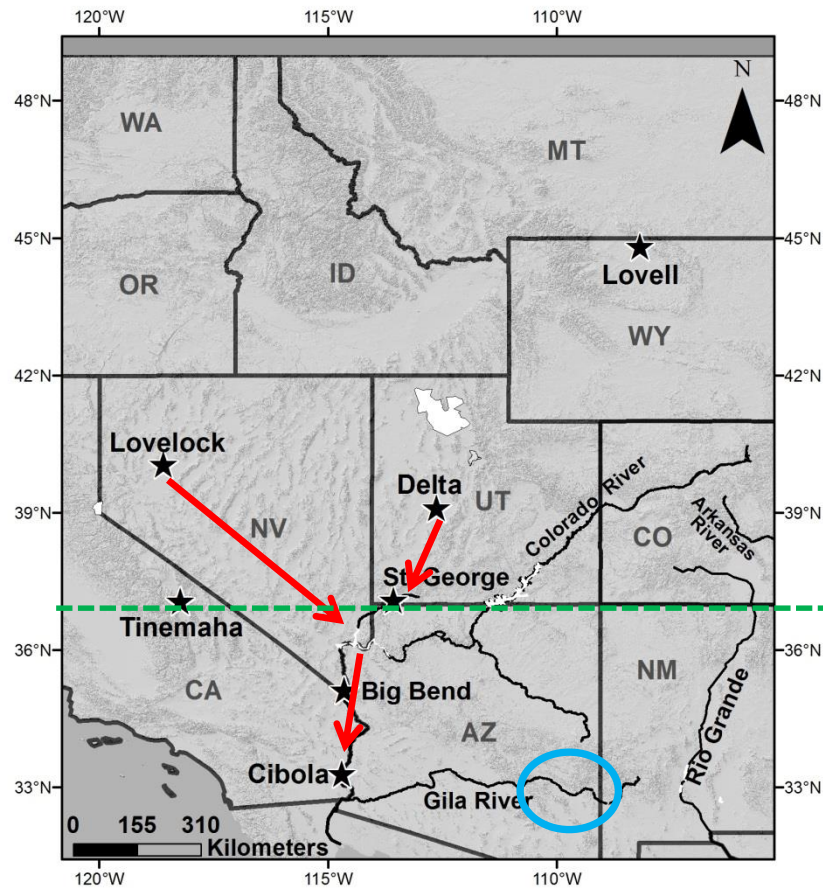
X

Diorhabda carinulata



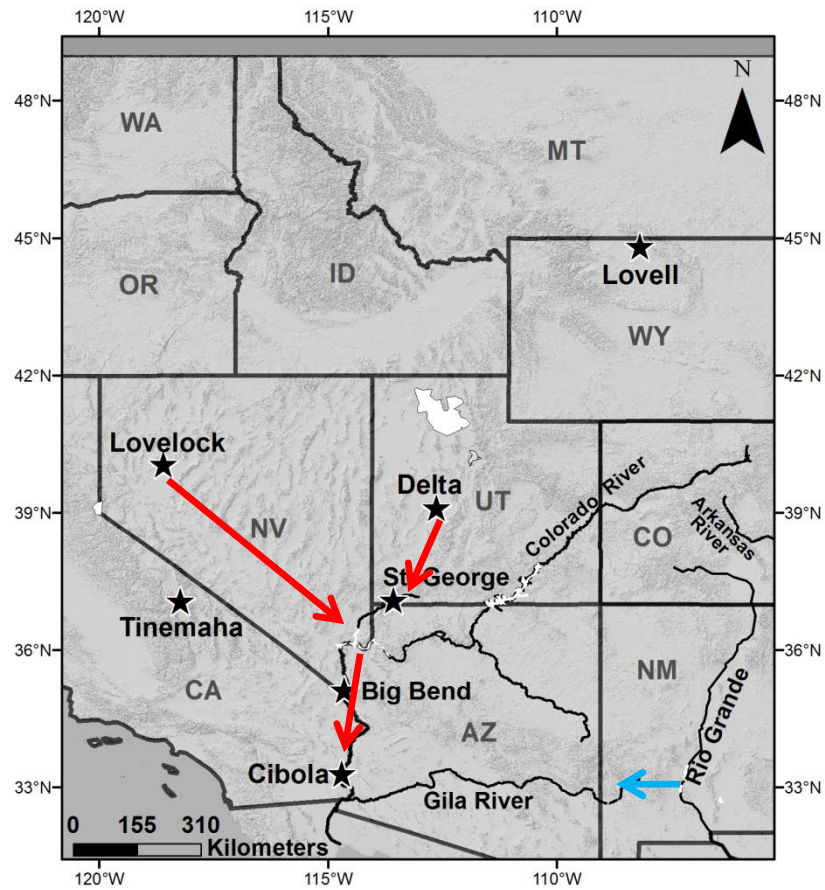
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← *D. carinulata*

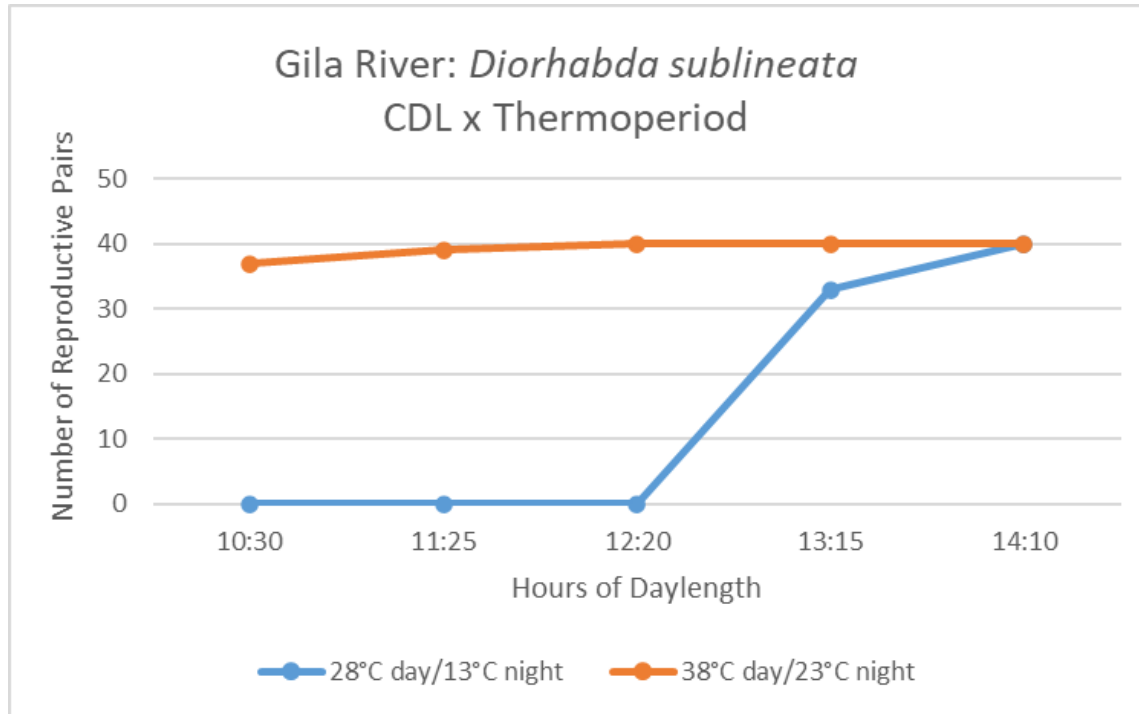
37th parallel



← *D. carinulata*

← *D. sublineata* and hybrids

The first experiment with Gila River beetles (subtropical tamarisk beetle) shows that photoperiodic diapause induction is highly sensitive to temperature, unlike with the northern tamarisk beetle. 40 pairs were tested per treatment



No diapause seen when daytime temperatures reach 38°C (100° F).

Thank you!

For questions please contact the authors. Alex Gaffke (alexander.gaffke@gmail.com), Tom Dudley (tdudley@msi.ucsb.edu) and Dan Bean (dan.bean@state.co.us)

We would like to thank a long list of researchers and cooperators at MSU, UC Santa Barbara and the Colorado Department of Agriculture as well as a number of cooperating institutions. All work presented was a team effort.

